

## *In vitro* Anti-inflammatory Effects of Thai Herb Essential Oils ฤทธิ์ต้านอักเสบของน้ำมันหอมระเหยจากสมุนไพรไทยในหลอดทดลอง

Nancharee Homnan (นันทิ้จารี หอมนาน)\* Dr. Chanphen Wiwat (คร.จันทร์เพ็ญ วิวัฒน์)\*\*

Dr.Suchitra Thongpraditchote (คร.สุจิตรา ทองประคิษฐ์ โชต์)\*\*\* Dr. Krit Thirapanmethee (คร.กฤษณ์ ถิรพันธุ์เมธี)\*\*\*\*

#### ABSTRACT

Thai herb essential oils have been used in traditional medicine for a long time. Several studies have reported a variety of its actions such as antibacterial, antifungal, antioxidant, anticancer and cytotoxic activities. In addition, many reports showed some essential oils also have anti-inflammatory effects in murine cell lines, however, the reports in human cell have been limited. Therefore, the goal of this study was to evaluate the anti-inflammatory activities of 10 Thai herb essential oils on lipopolysaccharide (LPS) induced inflammation in phorbol 12-myristate 13-acetate (PMA) differentiated THP-1 macrophage. THP-1 was pretreated with Thai herb essential oils for 1 hour prior to LPS. Then, the cells were harvested and subject to assay for cyclooxygenase 2 (COX-2) activity and cuture medium were collected to estimate prostaglandin  $E_2$  (PGE<sub>2</sub>) level. Among ten species of essential oils, Lesser galanga ([*Boesenbergia pandurata* (Roxb.)Schltr] oil showed the excellent activity to inhibit COX-2 production and PGE<sub>2</sub> with 100% and 93.3%, respectively. The COX-2 IC<sub>50</sub> of tested oil was 51.2 µg/ml. Thus, the results suggested that Lesser galanga essential oils are potential for inflammatory condition and may compose of several active compounds with anti-inflammatory properties. Further experiments are required to deeply analysis of their active compounds, mechanism of actions and application into medicinal use for anti-inflammatory purpose.

#### บทคัดย่อ

น้ำมันหอมระเหขจากสมุนไพรไทขมีการนำมาใช้ในขาแผนโบราณมาเป็นเวลานาและมีงานวิจัยหลายเรื่องที่ ราขงาน ฤทธิ์ของน้ำมันหอมระเหขาหล่านี้ เช่น ฤทธิ์ด้านเชื้อแบคทีเรียและเชื้อรา ฤทธิ์ด้านอนุมูลอิสระ และฤทธิ์เป็นพิษต่อเซลล์ นอกจากนี้ยังมีราขงานจำนวนมากแสดงให้เห็นว่าน้ำมันหอมระเหขบางชนิดมีฤทธิ์ในการด้านการอักเสบอีกด้วย แต่ส่วน ใหญ่มักเป็นการทดลองที่ทำในเซลล์ของหนู ส่วนการทดลองในเซลล์มนุษย์นั้นมีน้อยมาก ดังนั้นการทดลองนี้จึงมีเป้าหมาย ที่จะประเมินฤทธิ์ด้านการอักเสบของน้ำมันหอมระเหขากสมุนไพรไทยทั้ง 10 ชนิด โดยใช้เซลล์ THP-1 ที่ถูกกระตุ้นให้เกิด การอักเสบด้วยสาร lipopolysaccharide (LPS) หลังจากนั้นจะเก็บเซลล์เพื่อทดสอบการทำงานของเอนไซม์ cyclooxygenase-2 (COX-2) และเก็บอาหารเลี้ยงเซลล์เพื่อวัดปริมาณ prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) จากผลการทดลองพบว่าน้ำมันหอมจากกระชาย [*Boesenbergia pandurata* (Roxb.) Schltr] ให้ผลยับยั้งเอนไซม์ COX-2 ได้ดีที่สุด โดยขับยั้งกิจกรรมของเอนไซม์ COX-2 ได้ 100% ยังยั้งสาร PGE<sub>2</sub> ได้ 93.3% และความเข้มข้นค่ำสุดที่สามารถชับยั้งเอนไซม์ (COX-2) ได้ 50% (inhibitory concentration 50, IC<sub>so</sub>) ก็อ 51.2 µg/ml ผลการทดลองนี้จิ้ให้เห็นว่าน้ำมันหอมระเหยจากกระหาย มีฤทธิ์ที่น่าสนใจในการด้านอักเสบและมี สารสำคัญหลายชนิดที่มีคุณสมบัติด้านการอักเสบ โดยขณะนี้กำลังอยู่ระหว่างการศึกษาวิเคราะห์สารสำคัญ กลไกการออก ฤทธิ์ เพื่อประชุกต์ใช้เป็นยาสำหรับด้านกรอักเสบต์อิกเสบคด

Key Words : Essential oils, COX-2, PGE<sub>2</sub>

<mark>คำสำคัญ:</mark> น้ำมันหอมระเหย เอนไซม์ไซ โคลออกซิจิเนสทู โพรสตาแกลนดินอีทู

\*\* Associate Professor, Department of Microbiology, Faculty of Pharmacy, Mahidol University

<sup>\*</sup> Student, Master of Science Program in Biopharmaceutical Science, Department of Microbiology, Faculty of Pharmacy, Mahidol University

<sup>\*\*\*</sup>Associate Professor, Department of Physiology, Faculty of Pharmacy, Mahidol University

<sup>\*\*\*\*</sup>Lecturer, Department of Microbiology, Faculty of Pharmacy, Mahidol University



#### Introduction

Inflammation is a common mechanism of the immune system that protect against injuries caused by physical wounds and non-self-invaders. The inflammation response is vascular changes and white blood cell recruitment at the site which has been invaded or can be stimulated throughout the body. Although, the purpose of the inflammatory process is to eradicate the invaders and start repairing process to maintain general physiological functions, but long-term overinflammation might cause tissue damages and associated with many diseases such as rheumatoid arthritis, inflammatory bowel disorders, Alzheimer's disease and also cause of various physical dysfunctions. During inflammation, white blood cells secrete many inflammatory cytokines (interleukin-1 $\beta$  [IL-1 $\beta$ ], interleukin-6 [IL-6] and tumor necrosis factor- $\alpha$  $[TNF-\alpha]$ ) and involved inflammatory mediators such as nitric oxide (NO) and prostaglandin  $E_2$  (PGE<sub>2</sub>), which are generated by inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 (COX-2) respectively (Kim et al.,2013). Nowadays, numerous evidences have pointed out that cyclooxygenase (COX) is main enzyme that plays the important role in chronic inflammatory diseases. It exists in two isoforms, COX-1 and COX-2. Normally, COX-1 is responsible for protecting the gastric mucosa and maintaining homeostasis while COX-2 is playing a crucial part in the inflammatory process (Abdel-Azeem et al., 2009 and El-Sayed AAA et al.,2012). The role of COX-2 in driving inflammation is converts arachidonic acid (AA) that release from membrane phospholipid by phospholipase A2 (PLA2) into prostaglandin H<sub>2</sub> (PGH<sub>2</sub>) and finally prostaglandin E synthase (PGES) metabolite into PGE, which is mediator that related to the classic signs of inflammation: redness, warmth, swelling and pain (Ricciotti E et al., 2011). PGE2 act as vasodilators to promote vascular permeability and increased blood flow into the inflamed tissue leading to redness, swelling and edema. Furthermore, warmth and pain result from the action of  $PGE_2$  stimulates peripheral sensory neurons and the preoptic area of the hypothalamus to promote pyrogenic effects. Therefore, this study is focusing on COX-2 and  $PGE_2$  which would be beneficial in the treatment of pain and inflammation without interfering physiological processes.

There are several reports about anti-inflammatory effect of Thai herbs, such as allylpyrocatechol (APC) isolated from Piper betle Linn. (Betel) in LPSstimulate Raw 264.7. The APC was inhibited NO and PGE<sub>2</sub> production in a dose dependent manner as also reduced iNOS, COX-2, IL-12p40 and TNF-Ω mRNA expression (Sarkar et al., 2008). Boesenbergia pandurata (Roxb.) Schltr (Lesser galanga) extracts inhibited LPS induction of PGE2 in RAW 264.7 cell lines. Panduratin A and hydroxypanduratin A showed strong activities against PGE<sub>2</sub> release, with IC<sub>50</sub> values of 10.5 and 12.3  $\mu$ M, respectively, and moderate effects on TNF- $\alpha$  (IC<sub>50</sub> = 60.3 and 57.3 µM, respectively (Tewtrakul et al., 2009). The different constituents of Zingiber cassumunar Roxb. (Plai) markedly decreased COX-2 and PGE<sub>2</sub> production in LPS-stimulated human dental pulp cells (Aupaphong et al.,2013). Moreover, phenylbutenoid dimers 1 and 2 that isolated from the rhizomes of Zingiber cassumunar Roxb. demonstrated potent COX-2 inhibitiory activity with the IC50 values of 2.71 and 3.64 µM, respectively in LPS induced RAW 264.7 (Han et al., 2005).

There are several studies have reported various anti-inflammatory effect of essential oil, however, the reports on actions of Thai herb essential oil on the inflammatory activity in human cells have been limited. In this study, 10 Thai herb essential oils were evaluated for their anti-inflammatory functions on human monocytic cell line (THP-1). The cyclooxygenase-2 activities was examined using the enzymatic assay kit and the prostaglandin  $E_2$  was determined by enzyme immunoassay.

#### Methodology

#### Chemicals

Roswell Park Memorial Institute (RPMI) 1640 medium (Gibco, Karlsruhe, Germany), Fetal Bovine Serum (FBS; PAA, Cölbe, Germany), NaHCO<sub>3</sub>, Penicillin/Streptomycin solution were obtained from GIBCO (Grand Island, NY). Phosphate-buffered saline (PBS), trypan blue, Phorbol 12-myristate 13-acetate (PMA; Sigma, St. Louis, MO, USA), lipopolysaccharide ( LPS) from *Escherichia coli* 055:B5,  $\gamma$ -irradiated ( Sigma, St. Louis, MO, USA) , DMSO, Celicoxib (Sigma, St. Louis, MO, USA), COX Activity Assay Kit and PGE<sub>2</sub> Enzyme Immunoassay kit were purchased from Cayman Chemical (Ann Arbor, MI, USA).

#### Test compounds

Ten essential oils which were obtained from Holy basil leaves and flowering shrub (*Ocimum tenuiflorum* L.), Hairy basil leaves (*Ocimum americanum* L.), Sweet basil leaves and flowering shrubs (*Ocimum basilicum* L.), Betel leaves (*Piper betle* L.), Guava leaf (*Psidium guajava* L.), Kaffir lime leaves (*Citrus hystrix* DC.), Lemongrass leaves (*Cymbopogon citratus* (DC.) Stapf.), Galanga rhizomes (*Alpinia galangal* (L.) Wild.), Lesser galanga rhizomes [*Boesenbergia pandurata* (Roxb.) Schltr.] and Plai rhizomes (*Zingiber cassumunar* Roxb.) were purchased from Thai-China Flavours and Fragrances Industry Co., Ltd.

#### THP-1 culture, treatment and stimulation

THP-1 human monocytes was cultured in RPMI 1640 medium supplemented with 10% fetal bovine serum and penicillin/streptomycin at 37 °C and 5%  $CO_2$  in a humidified incubator. Cells will be pelleted *via* centrifugation and assessed for viability using the Trypan-blue exclusion method. Viable cells were plated at a density of  $1 \times 10^7$  cells/mL in 100 mm culture dish. To differentiate monocytes into macrophages, THP-1 cells were treated with 40 µg/mL of PMA for 48 hours. Followed by adding each essential oil at desired concentration and further incubated for 1 hour. The inflammation will be induced by adding 1 µg/mL of LPS and incubate for 24 hours. After incubation, the cells were harvested in Tris-HCl pH 7.8 containing 1 mM EDTA by cell scraper then centrifugation for further analysis of COX-2 activity assay. All supernatants will be kept at -80 °C for PGE<sub>2</sub> determination.

# Assay of COX-2 Activity by COX Activity Assay kit

To analyze the mediation of COX-2, THP-1 cells were lysed in cold lysate buffer to release endogenous COX enzyme. Then, COX-2 level were measured with COX Activity Assay kit from Cayman Chemical (Ann Arbor, MI, USA). The measurement was carried out according to manufacturer's manual. The COX activity assay kit will be detected the colorimetric of oxidized N,N,N',N'-tetramethyl-p-phenylenediamine (TMPD) at 590 nm that produced by peroxidase activity of COX. The COX-1 inhibitor (SC-560) will be added to subtract absorbance from total COX activity. After that, calculate the IC<sub>50</sub> using Graph Pad prism software (Graph Pad Inc, San Diego, USA) with Equation: log concentration (inhibitor) vs. normalized response --Variable slope.

### Assay of Prostaglandin E<sub>2</sub> by Enzyme Immunoassay kit

PGE<sub>2</sub> level in cultured medium was evaluated by enzyme Immunoassay kit from Cayman Chemical



(Ann Arbor, MI, USA). The measurement according to the manufacturer's instructions.  $PGE_2$  in the supernatants ( $PGE_2$  Tracer) in constant concentration were competitive binding to  $PGE_2$  monoclonal antibody that coated the bottom of the well. The plate will be washed to remove all unbound reagents, and added Ellman's reagent (which contains the substrate to AChE) each well for color development. Then, determine the intensity between 405 and 420 nm. The effects of 10 Thai herb essential oils on  $PGE_2$  productions were calculated by comparison with a standard curve and represent as  $PGE_2$  concentration. Finally,  $PGE_2$  level be calculated and reported as percent inhibition.

#### Results

In this study, we first measured the effect of 10 Thai herb essential oils on COX-2 activity. THP-1 were supplemented with culture medium with or without 100  $\mu$ g/ml of 10 Thai herb essential oils for 1 hour and activated with 1  $\mu$ g/mL of LPS to stimulated COX-2 for 24 hour. The result showed that Lesser of each sample and PGE2-acetylcholinesterase conjugate

galanga essential oil exhibited the best COX-2 inhibitory effect with 100% inhibition whereas guava leaf and lemongrass gave 54.6% and 50.5%, respectively (Table 1).

After that, culture medium were collected for evaluate  $PGE_2$  level whether perform with  $PGE_2$ Enzyme Immunoassay kit. As shown in Table 1, percentage inhibition of  $PGE_2$  in the conditioned media treated with Betel vine essential oil showed the most potent inhibitor of  $PGE_2$  production with 99.9% while Holy basil, Plai and Lesser galanga displayed 99%, 94.8% and 93.3%, respectively. From the result of COX-2 and  $PGE_2$  inhibition, Lesser galanga essential oils demonstrated the most interesting percentage inhibition. Therefore, The 50% inhibition concentration ( $IC_{50}$ ) of this essential oil was investigated. The  $IC_{50}$  of Lesser galanga essential oil was 51.2 µg/ml (Figure 1).

Essential oil	% COX-2 inhibition	% PGE <sub>2</sub> inhibition
DMSO	0	0
Celecoxib	45.5	90.7
Kaffir lime	0	69.2
Plai	18.2	94.8
Galanga	0	90.1
Holy basil	40	99
Hairy basil	18.2	87.8
Guava	54.5	60.7
Lesser galanga	100	93.3
Betel	26.3	99.9
Sweet basil	21.9	0
Lemongrass	50.4	93.3

Table 1 Effect of 10 Thai herb essential oils and their inhibitory effect of COX-2 and PGE<sub>2</sub> level on LPS-activated THP-1 cells.





Figure 1 Effect of Lesser galanga essential oils on COX-2 enzyme activity on LPS-activated THP-1 cells. To derive IC50, THP-1 macrophages were incubated with different concentration of Lesser galanga essential oil for 1 hour. Then stimulated COX-2 using 1 µg/mL of LPS for 24 hour. Cell were collected and measured COX-2 activity by COX Activity Assay kit from Cayman Chemical

From the result of COX-2 and PGE<sub>2</sub> inhibition, Lesser galanga essential oils demonstrated the most interesting percentage inhibition. Therefore, The 50% inhibition concentration (IC<sub>50</sub>) of this essential oil was investigated. The IC<sub>50</sub> of Lesser galanga essential oil was 51.2  $\mu$ g/ml (Figure 1). Therefore, this results indicates the capacity of Lesser galanga essential oil to inhibit COX-2 activity of THP-1 cell.

#### **Discussion and Conclusion**

Among of 10 Thai herb essential oils, Lesser galanga essential oil displayed the most promising effects with 100% COX-2 inhibition and 93.3%  $PGE_2$  inhibition. Moreover, Lesser galangal oil was inhibit COX-2 activity in dose-dependent manner with IC<sub>50</sub> value of 51.2 µg/ml on LPS stimulated THP-1. Therefore, we expected there were antiinflammatory ingredient in essential oil. Lesser galanga oils that analysed by GC and GC-MS showed the major constituents such as camphor, geraniol and (E)- $\beta$ -ocimene (Jantan *et al.*, 2001). The further studies will be performed on the pure compound, to find out the active compound, mechanism of actions, finally apply into medicinal use of anti-inflammatory purpose.

#### Reference

- Abdel-Azeem AZ, Abdel-Hafez AA, El-Karamany GS, Farag HH. Chlorzoxazone esters of some non-steroidal anti-inflammatory (NSAI) carboxylic acids as mutual prodrugs: Design, synthesis, pharmacological investigations and docking studies. Bioorg Med Chem 2009; 17(10):3665–70.
- Aupaphong V, Dechatiwongse Na Ayudhya T, Koontongkaew S. Inhibition of lipopolysaccharideinduced expression of cyclooxygenase-2 by *Zingiber cassumunar* Roxb. constituents in human dental pulp cells. ACAD J 2013; 7(33): 2451-8.



- El-Sayed MA, Abdel-Aziz NI, Abdel-Aziz AA, El-Azab AS, ElTahir KE. Synthesis, biological evaluation and molecular modeling study of pyrazole and pyrazoline derivatives as selective COX-2 inhibitors and anti-inflammatory agents. Bioorg Med Chem 2012; 20(10 Pt 2): 3306–16.
- Han AR, Kim MS, Jeong YH, Lee SK, Seo EK. Cyclooxygenase-2 Inhibitory Phenylbutenoids from the Rhizomes of *Zingiber cassumunar*. Chem Pharm Bull 2005; 53(11): 1466-68.
- Jantan IB, Basni I, Ahmad AS, Ali NAM, Ahmad AR, Ibrahim H. Constituents of the rhizome oils of *Boesenbergia pandurata* (Roxb.) Schlecht. from Malaysia, Indonesia and Thailand. Flavour Fragr J 2001; 16: 110–112
- Kim KN, Ko YJ, Yang HM, Hamb YM, Roha SW, Jeon YJ, et al. Anti-inflammatory effect of essential oil and its constituents from fingered citron (*Citrus medica* L. var. *sarcodactylis*) through blocking JNK, ERK and NF-KB signaling pathways in LPS-activated RAW 264.7 cells. Food Chem Toxicol 2013; 57:126– 31.

- Ricciotti E, FitzGerald AG. Prostaglandins and inflammation. Arterioscler Thromb Vasc Biol 2011; 31(5):986–1000.
- Sarkar D, Saha P, Gamre S, Bhattacharjee S, Hariharan C, Ganguly S, et al. Antiinflammatory effect of allylpyrocatechol in LPS-induced macrophages is mediated by suppression of iNOS and COX-2 via the NF-KB pathway. Int Immunopharmacol 2008; 8: 1264–71.
- Tewtrakul S, Subhadhirasakul S, Karalai C, Ponglimanont C, Cheenpracha S. Anti-inflammatory effects of compounds from *Kaempferia parviflora* and *Boesenbergia pandurata*. Food Chem 2009; 115: 534–38