



Insecticidal Activities of *Piper retrofractum* Extracts Against *Aedes aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae)

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ABSTRACT

Mosquito borne diseases such as dengue fever, filarial are endemic disease of more than hundred countries worldwide. It causes millions of morbidity and mortality of people. Vector control by using chemical insecticides is one important approach to reduce the infected vectors. Plant derived insecticide is an alternative to control by chemical insecticide. In this study was to investigate adulticide and larvicide properties of *Piper retrofractum*. Essential oil was extracted from a mixture of fresh ground fruit and water by hydrodistillation. Adulticide activities were determined by applying drops of essential oil in acetone to the thoraces of mosquitoes. The results showed an LD_{50} of $8.86\% \pm 0.69$ and an LD_{50} of $19.16\% \pm 2.3$ for adult *Aedes aegypti* and an LD_{50} of $6.95\% \pm 0.51$ and an LD_{99} of $14.44\% \pm 2.49$ for adult *Culex quinquefasciatus*. Crude extracts were prepared by macerating dried fruit with 3 organic solvent, methanol, ethyl acetate, hexane and aqueous. Larvicide activities of the diluted crude extracts were diluted to the test concentrations with in acetone, and the crude aqueous extract with distilled water. The methanol extract showed the highest larvicidal activity for *Ae. aegypti*, the LC₅₀ being 2.2 ppm. The LC₅₀s for *Cx. quinquefasciatus* were similar, being 2.1, 2.4, 2.9, and 431.3 ppm for the methanol, ethyl acetate, hexane, and aqueous extracts, respectively.

Keywords: Piperaceae, Piper retrofractum, Larvicide, Adulticide, Mosquito control

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Introduction

Mosquito control strategies are generally focused on controlling mosquitoes in either the adult stage or the larval stages of the life cycle. Various physical, biological, and chemical methods have been used to control mosquito vectors, and the method used in a particular area will depend on the specific situation in that area and the budget available. Chemical control methods are popular and widely used in many countries because they are easy to perform and effective. Paul Hermann Muller discovered DDT in 1874 (WHO, 1979). DDT has been widely used to control a range of insect pests, including insect vectors, but DDT use was restricted in 2001 because of the serious negative impacts it has had on the environment. The regular and widespread use of insecticides has resulted in insect vectors increasingly developing resistance to insecticides (Gakhar et al., 2013). Temephos, an organophosphate pesticide, is commonly coated onto the surfaces of sand particles that are then spread in campaigns to control dengue (CDC, 2014). Temephos is very effective at killing mosquito larvae, but its unpleasant smell, tendency to cause films to form on water surfaces, and concern about the health risks it poses to humans through its presence in drinking water mean that the general public may not easily accept the use of temephos to control insect vectors. There is currently great interest in studying the insecticidal properties of plants. Insecticides extracted from plants can be used as alternatives to synthetic insecticides for controlling insect vectors. Tropical plants have great biodiversity, meaning that there is the strong possibility that some plants can be used to develop new insecticides.

The Piperaceae family contains plants with spicy flavors. The family contains roughly 3,920 species in 13 genera. The two main genera in the Piperaceae family are the Piper genus, which contains 2,000 species, and the Peperomia genus, which contains 1,600 species. Piperaceae plants are commonly found in South East Asia, and many have long been used in cooking and traditional medicines (Chaveerach et al., 2006). The major chemical constituents of the essential oils of Piperaceae plants beta-sitosterol, cineole, terpinan-4-ol, are 1-betacaryophyllene, piperodione, sitisterol, and venerol (Banerji, 2002; Kubo, 2013).

Many of the plants in the Piperaceae family have adulticidal and/or larvicidal activities toward insects. For example, ethanol extracts of Piper betle can kill the larvae of Aedes aegypti, with an LC₅₀ of 177.6 ppm (Komalamisra et al., 2005). Ethanol extracts of Piper longum, Piper ribesoides, Piper sarmentosum, and Piper nigrum have been found to have LC₅₀ concentrations of 2.2, 4.0, 8.1, and 0.9 ppm, respectively, toward Ae. aegypti larvae (Chaithong et al., 2006; Simas et al., 2007). Hexane extracts of P. longum, P. nigrum (white pepper), and P. nigrum (black pepper) have been found to have LC_{50} concentrations of 0.017, 0.024, and 0.007 ppm, respectively, toward Ae. aegypti larvae (Kumar et al., 2010). The essential oil of the fruit of P. longum has been found to have LC₅₀ concentrations, in solution, of 6.21 and 6.35 µg.mg⁻¹ toward females of a field strain and a laboratory strain, respectively, of Ae. aegypti (Chaiyasit et al., 2006). In the study presented here we investigated the adulticidal activity of the essential oil of Piper retrofractum fruit and the larvicidal activities of crude extracts of the fruits toward culicine mosquito vectors.



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Objective of the study

To evaluate adulticide and larvicide activity of *P. retrofractum* extracts against *Ae. aegypti* and *Cx. quinquefasciatus*.

Material and Methods

Plant material

Mature *P. retrofractum* fruits were collected from three houses in Huay-sathon village, Tha-mi District, Chanthaburi Province, Thailand, in April and May 2014. The plants and fruits had not been treated with synthetic insecticides because the owners grew the plants to use the fruits in their own kitchens. Specimens of the plants were identified by Dr. Chalermphol Suwanphakdee from Kasetsart University, Thailand and the specimens were archived at the Queen Sirikit Botanical Garden, Mae-rim District, Chiangmai Province, Thailand as voucher specimen QBG No.79271.

Essential oil preparation

The essential oils were extracted by hydro distillation method. Each fruit was approximately 5 cm long and 1 cm in diameter. A 1 kg sample of the fresh *P. retrofractum* fruits was washed with tap water, air dried, then ground in a blender. The ground fruit and 2 L of distilled water were added to a distillation apparatus, and the essential oil was extracted by distilling the mixture for 3 hours. The product was kept at 4 °C until use.

Preparation of crude extracts

The crude extracts were extracted by maceration methods with 4 different solvents as methanol, ethyl acetate, hexane or aqueous. Four 10 g samples of the dried fruits were crushed in a blender and then each sample was macerated with 500 mL of a solvent for 1 day. Each mixture was then filtered

and the solution was concentrated (using a rotary evaporator for the solutions in organic solvents and by lyophilization for the aqueous solution). The crude extracts were kept at 4 °C until use.

Mosquito strains

Susceptible strain of Ae. aegypti and Cx. quinquefasciatus were provided by the Insecticide Research Unit, Department of Medical Entomology, Faculty of Tropical Medicine, Mahidol University, Bangkok, Thailand. The rearing procedure involved putting the eggs in a 25 cm \times 50 cm \times 10 cm tray containing 700 mL of dechlorinated tap water. Twenty-four hours after the eggs had hatched, 200 late first instar larvae were transferred to a new tray to be reared, to avoid the larvae becoming overcrowded. The larvae were provided with Hikari® Lionhead fishfood pellets (Kyorin[®];Himeji, Japan). Third instar larvae were selected for the larvicidal activity tests. The remaining larvae continued to be reared to the adult phase. The adults were maintained at 28±2 °C until they were used in an experiment, and they were provided with 10% sugar solution soaked cotton

Adulticidal activity

The adulticidal activity against *Ae. aegypti* and *Cx. quinquefasciatus* was determined by topically applying a solution of the essential oil in acetone on pronotum of adult *Ae. aegypti* and *Cx. quinquefasciatus* followed a protocol described by the World Health Organization (2006) with slight modifications. The working solutions of essential oil in acetone were prepared in 5%, 7%, 9%, 11%, 13%, and 15%. Fifty female mosquitoes in each species, non-blood fed, 2-5 days old were used in each concentration. It was anesthetized with ether and then placed on a cold plate before tested. A 0.5 μ L drop of an essential oil in acetone was applied to the pronotum of each



mosquito using a Burkard[®] hand microapplicator (Burkard Scientific[®]; Uxbridge, United Kingdom). After exposure, the tested mosquitoes were transferred in to plastic cups provide with 10% sugar solution soaked cotton placed on covered net and held in appropriate condition for 24 hr. Damage or unrecovered mosquitoes were removed. Two control groups were used, the mosquitoes in one were exposed to acetone and the mosquitoes in the other were untreated. Three replicates of each test, using mosquitoes from separately reared batches, were performed.

Larvicidal activity

Larvicide activities were determined by diluted crude extracts in distilled water with third instar larvae using a method following World Health Organization guidelines (WHO, 2005) with slight modifications. Each extract was diluted to give the desired test concentration, and each test was performed on 100 third instar Ae. aegypti larvae and 100 third instar Cx. quinquefasciatus larvae. The concentration of organic solvent extract in acetone were 1.5, 2.0, 2.5, 3.0, 4.0, and 5.0 ppm and the water crude extract dissolved in distilled water were 150, 200, 400, 600, 800, and 1000 ppm. The mortality was determined after the larvae had been exposed for 24 hours. Three replicates of each test, using larvae from separately reared batches, were performed. The Median Lethal Dose (LD₅₀) and Median Lethal Concentration (LC₅₀) were calculated using Probit analysis (Finney, 1987).

Results

The essential oil extracted from the fresh *P. retrofractum* fruit was clear, had a sweet, spicy odor, and was soluble in acetone. A total of 1.25 mL

of essential oil was extracted from 1 kg of the fresh fruit by distillation (0.125% yield.). Exposure to 15% essential oil in acetone for 24 hrs was found to cause more than 90% mortality in both *Ae. aegypti* (92.7% mortality) and *Cx. quinquefasciatus* (98.7% mortality). No mortality occurred in either species in the acetone control and untreated control groups

The LD_{50} were 8.86% ± 0.69 for *Ae. aegypti* and 6.95% ± 0.51 for *Cx. quinquefasciatus*, as shown in Table 1.

Table 1 LD_{50} and LD_{95} values of P. retrofractum
essential oil against Ae. aegypti and
Cx. quinquefasciatus by topical application
after 24 h.

	Lethal Dose		
Species	(% of essential oil in acetone)		
	LD ₅₀ ±SE	LD ₉₅ ±SE	
Ae. aegypti	8.86 ± 0.69	19.16 ± 2.30	
Cx. quinquefasciatus	6.95 ± 0.51	14.44 ± 2.49	

The crude aqueous, methanol, and ethyl acetate extracts were yellow and contained particles, but the crude hexane extract was red and oily. The 10 g samples of dried *P. retrofractum* fruit gave crude extracts of 14% yield when methanol was used, 9% yield when ethyl acetate was used, 6% yield when hexane was used, and 20% when aqueous was used. The LC_{50} concentrations for *Ae. aegypti* larvae after 24 hours of exposure were 2.2, 2.9, and 2.7 ppm for the crude methanol, ethyl acetate, and hexane extracts, respectively, diluted in acetone, and 297.4 ppm for the aqueous extract diluted in distilled water as shown in Table 2. The LC_{50} concentrations for *Cx. quinquefasciatus* larvae after 24 hours of exposure



were 2.1, 2.4, and 2.9 ppm for the crude methanol, ethyl acetate, and hexane extracts, respectively, diluted in acetone, and 431.3 ppm for the aqueous extract diluted in distilled water as shown in Table 2.

Table 2LC50 values for Ae.aegypti and
Cx. quinquefasciatus larvae exposed
to methanol, ethyl acetate, hexane
and aqueous extracts of P. retrofractum
for 24 h.

	Lethal Concentration (ppm) \pm SE				
Solvent	Ae. aegypti		Cx. quinquefasiatus		
	LC ₅₀	LC ₉₅	LC ₅₀	LC ₉₅	
Methanol	$2.21 \pm$	$4.45~\pm$	$2.11 \pm$	$5.70 \pm$	
	0.28	1.13	0.4	0.45	
Ethyl	$2.96\pm$	$5.61 \pm$	$2.47 \pm$	$6.30 \pm$	
acetate	0.57	1.06	0.11	0.47	
Hexane	$2.77 \pm$	$5.79 \pm$	$2.90\pm$	$4.18\pm$	
	0.26	0.60	0.25	0.75	
Aqueous	$297.39\pm$	629.65	$431.28\pm$	$1,621 \pm$	
	0.78	± 1.43	1.12	1.73	

Discussion

The Piperaceae family contains 3,920 species, and at least 17 species have been found to have insecticidal properties (Marques and Kaplan, 2015). Piperidine alkaloids are active ingredients in many Piperaceae species (Mishra, 2010; Hieu *et al.*, 2014) but the concentrations of these alkaloids vary depending on the species, geographic area, stage of growth, and season (Bao *et al.*, 2014). Piperidine alkaloids can enter an insect through ingestion (Paula *et al.*, 2000) and the alkaloids can inhibit egg hatching (Tavares *et al.*, 2011) or act as adulticides (Rassami, 2011) or larvicides (Chansang *et al.*, 2005). The insecticidal activities of essential oil and crude extracts

of *P. retrofractum* fruits against *Ae. aegypti* and *Cx. quinquefasciatus* were determined in this study.

The estimated yield of the essential oil used in the adulticide tests was 0.125%, which is five times lower than the yield achieved using the same method from the closely related species P. longum (Choochote et al., 2006) The Cx. quinquefasciatus adults were affected more strongly by exposure to the essential oil than were the Ae. aegypti adults, the LD_{50} being 6.9% for Cx. quinquefasciatus and 8.8% for Ae. *aegypti* as is shown in Table 1. Seizures occurred in the mosquitoes that had been exposed to the essential oil, once they had recovered from the anesthetic, some of the mosquitoes lost control of and then actually lost their legs and wings. These effects continued for 1 hour, then some of the mosquitoes died. Essential oil extracted from P. longum had LD_{50} of 6.21 and 6.35 µg.mg⁻¹ against laboratory strain and field strain of adult Ae. aegypti females in a previous study by Choochote et al. (2006). P. longum appears to have a stronger adulticidal activity than did P. retrofractum (which is closely related) in our study the LD₅₀ of *P. retrofractum* essential oil was 8.8%, which is equivalent to 16.56 μ g. mg⁻¹. Essential oil extracted from P. retrofractum will therefore be suitable for controlling the polluted breeding species Cx. quinquefasciatus. Essential oil extracted from P. retrofractum has been used in shampoo for treating Pedicular capitis in humans (Rassami, 2011). Further study is required for using P. retrofractum essential oil to control Rhipicephalus sanguineus and Ctenocephalides canis in pets.

The LC_{50} concentrations for the organic solvent extracts were not statistically different from each other, but the LC_{50} for the aqueous extract was much higher. The *Ae. aegypti* larvae were 1.5 times



more sensitive than the Cx. quinquefasciatus larvae to the aqueous extract. The methanol extract had the strongest larvicidal activity for both Ae. aegypti and *Cx. quinquefasciatus*. The methanol extracts LC_{50} for Ae. aegypti larvae was 142 times lower than the LC_{50} of aqueous extract. The methanol extract LC₅₀ for Cx. quinquefasciatus was 188 times lower than the LC_{50} of aqueous extract. The larvae responded to the crude extracts within four hours. After that time they moved less than before, became unable to swim to the water surface, and some eventually died. The modes of action the crude extracts was interesting, and need further studies. The larvicidal activities of species in the Piperaceae family have been reported many times. For example, an ethanol extract of *P*. betle had an LC_{50} of 177.62 ppm against Ae. aegypti larvae (Komalamisra et al., 2005), an ethanol extract of P. longum was found to have an LC50 of 2.23 ppm against Ae. aegypti larvae (Chaithong et al., 2006) and also ethanol extracts of white and black P. nigrum were found to have LC₅₀ concentrations of 0.356 and 405 ppm, respectively (Kumar et al., 2010). The results of our study agreed with the results of a study by Chansang et al. (2005) who found that crude extracts of dried ripe P. retrofractum fruit were larvicides for Ae. aegypti and Cx. quinquefasciatus with LC_{50} of 79 and 135 ppm, respectively. The synthesis of the compounds responsible for the insecticidal activities of the P. retrofractum extracts could possibly be developed to new insecticides for mosquito control. However, the crude extracts should be tested against other mosquito species, such as Ae. albopictus, Cx. tritaeniorhynchus, and Anopheles spp., including other agricultural insects. Since it is the editable plant and the ease preparation of the crude extract by maceration made it can apply in the field to control the pest insects easily. It is eco-friendly and can be the alternative to the chemical synthetic insecticide.

Conclusion

Essential oil extracted from *P. retrofractum* fruit was an adulticide for *Ae. aegypti* and *Cx. quinquefasciatus*, the LD₅₀ values being 8.8% for *Ae. aegypti* and 6.9% for *Cx. quinquefasciatus*. The crude methanol extract of dried *P. retrofractum* fruits was a stronger larvicide for *Ae. aegypti* and *Cx. quinquefasciatus* than that were the other crude extracts. A methanol extract of *P. retrofractum* could potentially, therefore, be developed for use in mosquito control programs in urban and rural areas.

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References

- Banerji A, Sarkar M, Datta R, Sengupta P, Abraham K. Amide from *Piper brachystachyum* and *Piper retrofractum*. Phytochem Lett 2005; 59: 897-901.
- Bao N, Ochir S, Sun Z. Occurrence of piperidine alkaloids in Piper species collected in different areas. J Nat Med 2014; 68:211-4.



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Temephos. [online] 2014 [cited 2014 Jan 30]. Available from:http://www.cdc.gov/niosh/ pel88/3383-96.html/

Centers for Disease Control and Prevention:

- Chaithong U, Choochote W, Kamsuk K, Jitpak A,
 Tippawangkosol P, Chaiyasit D, Champakaew
 D, Tuetun B, Pitasawat B. Larvicidal effect of
 pepper plants on *Aedesaegypti* (L.) (Diptera:
 Culicidae). J Vector Ecol 2006; 31:138-144.
- Chaiyasit D, Choochote W, Rattanachanpichai E,
 Chaithong U, Chaiwong P, Jitpakdi A,
 Tippawangkosol P, Riyong D, Pitasawat B.
 Essential oils as potential adulticides against
 two populations of *Aedes aegypti*, the
 laboratory and natural field strains, in Chiang
 Mai province, northern Thailand. Parasitol
 Ref 2006; (6)99: 715 721.
- Chansang U, Zahiri S, Bansiddhi J, Boonruad T,
 Thongsrirak P, Mingmuang J. Mosquito
 larvicidal activity of aqueous extracts of long
 pepper (*Piper retrofractum*) from Thailand.
 J Vector Ecol 2005; 30:195-200.
- Chaveerach A, Sudmoon R, Tanee T, Mokkamul P. Three new species of Piperaceae from Thailand. Acta Phytotaxon Sin 2006; 44: 447-453.
- Choochote W, Chaithong U, Kamsuk K, Rattanachanpichai E, Jitpakdi A, Tippawangkosol P. Adulticidal activity against *Stegomyiaaegypti* (Diptera: Culicidae) of three Piper spp. Rev Inst Med Trop Sao Paulo 2006; 48: 33-37.
- Finney D. Probit analysis 3rdedn. Cambridge University Press 1971.

- Gakhar SK, Sharma R, Sharma A. Population genetic structure of malaria vector *Anopheles stephensi* Liston (Diptera: Culicidae). Indian J Exp Biol 2013; 51: 273 - 279.
- Hieu D, Thang D, Hoi M, Ogunwande A. Chemical composition of Essential Oil from Four Vietnamese Species of Piper (Piperaceae). J Oleo Sci 2014; 63: 211-217.
- Komalamisra N, Trongtokit Y, Rongsriyam Y, Apiwathnasorn C. Screening for larvicidal activity in some Thai plants against four mosquito vector species. Se Asian J Trop Med 2005; 36: 1412-1422.
- Kubo M, Ishii R, Ishino Y, Harada K, Matsui N, Akagi M, Kato E, Hosoda S, Fukuyama Y. Evaluation of Constituents of *Piper retrofractum* Fruits on Neurotrophic Activity. J Nat Prod 2013; 76: 769-773.
- Kumar S, Warikoo R, Wahab N. Larvicidal potential of ethanolic extracts of dried fruits of three species of peppercorns against different instars of an indian strain of dengue fever mosquito, *Aedes aegypti L.* (Diptera: Culicidae). Parasitol Res 2010; 107: 901-907.
- Marques M, Kaplan P. Active metabolism of the genus Piper against *Aedes aegypti*: natural alternative sources for dengue vector control. Unic Sci 2015; 20: 61-82.

Mishra P. Isolation, Spectroscopic characterization and computational modeling of chemical constituents of *Piper longum* natural product. J Ethnopharmacol 2010; 2: 78-86.

Paula VF, Barbosa C, Demuner, J, Pilo-Veloso D, Picanco C. Synthesis and insecticidal activity of new amide derivatives of piperine. Pest Manag Sci 2000; 56: 168-174.



- Rassami W, Soonwera M. Effect of herbal shampoo from long pepper fruit extracts to control human head louse of the Ladkrabang Children, Bangkok, Thailand. J Agi Tech 2011; 7: 331-338.
- Simas K, Lima EC, Kuster RM, Lage CL. 2007. Potential use of *Piper nigrum*ethanol extract against pyrethroid-resistant *Aedes aegypti* larvae. Rev Soc Bras Med Trop 2007; 40: 405-407.
- Tavares WS, Cruz I, Petacci F, Freitas SS, Serrao JE,
 Zanuncio JC. Insecticide activity of piperine: Toxicity to eggs of Spodoptera frugiperda (Lepidoptera:Noctuidae) and Diatraea saccharalis (Lepidoptera:Pyralidae) and phytotoxicity on several vegetables. J Med Plants Res 2011; 5: 5301-5306.

World Health Organization [online] 1979.

Environmental health criteria9: DDT and its derivative. [cited 2014 Jan 30] Available from: http://www.inchem.org/documents/ehc /ehc /ehc009.htm/

- World Health Organization [online] 2005. Guidelines for laboratory and field testing of mosquito larvicides. [cited 2014 Jan 30] Available from: http://apps.who.int/iris/bitstream/ 10665/69101/1/WHO_CDS_WHOPES_GCD PP_2005.13.pdf/
- World Health Organization [online] 2006. Guidelines for Testing Mosquito Adulticides for Indoor Residual Spraying and treatment of Mosquito Nets. [cited 2014 Jan 30] Available from: http://apps.who.int/iris/bitstream/10665/69296 /1/WHO_CDS_NTD_WHOPES_GCDPP_20 06.3_eng.pdf/